The rapid expansion of global trade and human travel has greatly accelerated the introduction of non-endogenous pathogens and pests.

The global climate change influences pathogen communities both directly and indirectly



Challenge 1



Difficulties in accurately determining species/population identity based on morphological traits->

Accurate determination of species/population identity is critical for risk assessment and the implementation of regulatory measures and appropriate disease control strategies.

Issues in Molecular Identification

- ✓ Molecular identification is not a panacea.
 - Your results may be as good as what you compared with (legacy errors and problems).
 - Different genes may tell different stories.
 - Molecular data without biological contexts have a limited value.
- √ What is species?
 - Biological, morphological, and phylogenetic species concepts.
 - They overlap but not perfectly.

Challenge 2

"There are known knowns. These are things we know that we know. There are known unknowns. That is to say, there are things that we know we don't know. But there are also unknown unknowns. There are things we don't know we don't know."

- Donald Rumsfeld -

HE PH WASH

Limited understanding of pathogen diversity in nature ->

There exist many "known knowns, known unknowns & unknown unknowns" threatening agricultural and ecological systems.

Challenge 3

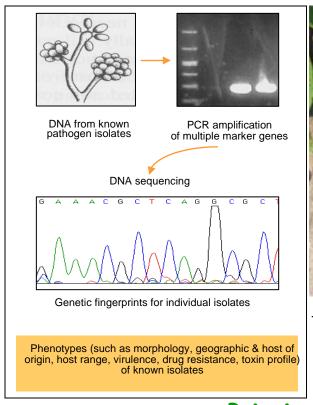


Limited efforts and mechanisms to support global cooperation and knowledge sharing->

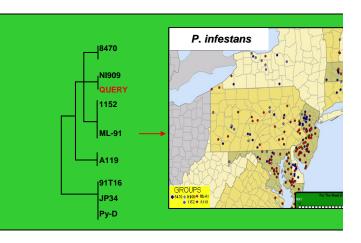
Mapping and documenting the diversity, distribution, and biology of major pathogens worldwide and sharing this information are essential to improve our ability to track and manage pathogens.

Phytophthora Database

Cyberinfrastructure supporting identification and monitoring of *Phytophthora* species



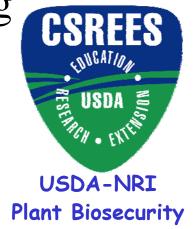




www.phytophthoradb.org

Principal Investigators

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- UC-Riverside: Mike Coffey
- North Carolina State University: Kelly Ivors
- USDA-ARS: Frank Martin & Kerry O'Donnell





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·PA Dept of Ag: S. H. Kim & E. Nikolaeva

·Seoul National University: Y. Lee, K. Jung & J. Park

·USDA-ARS SBML: A. Rossman, D. Farr & E. Cline

·APS Press: D. Erwin & O. Ribeiro









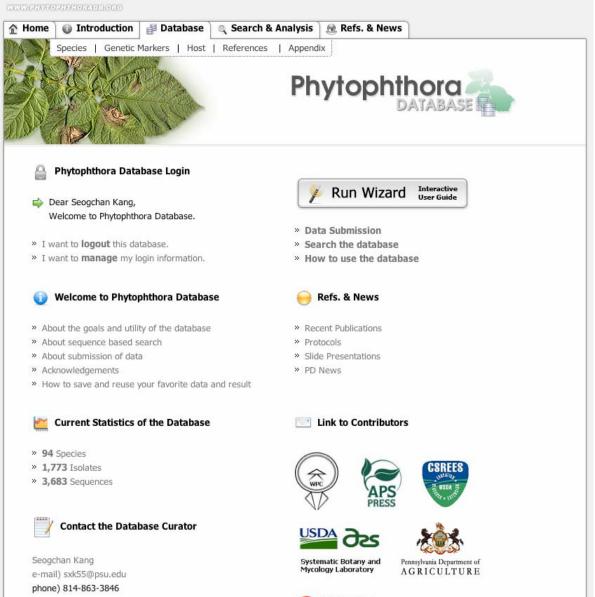
Specific Objectives of the PD Project

- ✓ To establish a comprehensive phylogenetic framework for the genus *Phytophthora*.
- To build a cyberinfrastructure that archives the genotypes, phenotypes, and distribution of individual *Phytophthora* species/isolates and provides data analysis and visualization tools.
- ✓ To develop and optimize molecular diagnostic tools for detecting and identifying *Phytophthora* species.

<u>Progress</u>

- ✓ Development of new phylogenetic markers based on the genome sequences of *P. ramorum*, *P. capsici*, *P. sojae* and *P. infestans*.
- ✓ Conducted a genus-wide phylogenetic analysis using representative species/isolates.
- ✓ Genotypic identification of ~3,000 isolates (>90 species).
- ✓ Establishment of the *Phytophthora* Database.
- ✓ Characterization of individual species complexes (e.g., P. cinnamomi, P. capsici)

Organization of the Pytophthora Database





Species Information Page

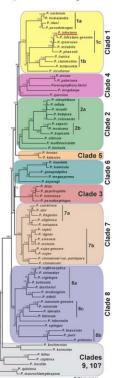


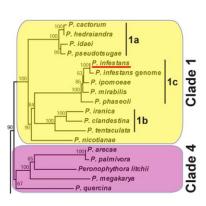


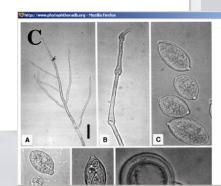
Phytophthora infestans

Phylogenetic Position within the Genus

This genus-wide phylogenetic tree contains 74 species, including Pythium vexans as the outgroup, and was built using sequences at seven loci (approximately 7700 nucleotides), including 605 Ribosomal Protein L10, Beta Tubulin, Enolase, Heat Shock Protein 90, Large Subunit rRNA, TigA gene fusion, and Translation Elongation Factor 1 alpha (Jaime Blair et al., unpublished data).











Phytophthora Infestans (Mont.) de Bary 1076 (Oomycetes, Pythlales) ■Botrytis infestans Mont. 1845

- =Peronospora infestans (Mont.) Casp. 1854
- = Botrytis fallax Desm. 1846
- = Peronospora fintelmannii Casp. 1052
- = Botrytis solani Harting 1846
- Phytophthora thalictri G.W. Wilson & Davis 1907
- = Phytophthora infestans f. sp. thalictri (G.W. Wilson & Davis) G.M. Waterhouse 1963
- = Peronospora trifurcata Unger 1947

Notes: The original name applied to this comycete was Gangraena tuberum solani, by Martius in 1042. After a period of debate (see Erwin 8 Ribeiro 1996), it was described by Montagne in 1845 and given the name Bottytis Infestans. It has received much renown as the cause of the great Irish potato famine of the 1840s, and it is the type of the genus *Phytophthora*. The name *Bothythis* devastatrix (alternate spelling vastatrix or devastrix) i.b. 1845 was published previous to Bothytis infestans, and has been listed as a synonym by various authors (e.g. Waterhouse 1963, Erwin & Ribeiro 1996). If this were true, the epithet vastatrix would have priority for this species. Waterhouse (1970) retained the name *Phytophthora infestans*, arguing incorrectly that *Botryts vastatrix* was an invalid name because it was published in a newspaper, but this is permitted by the Code previous to 1953 (Art. 30.3). While it is in fact valid, Botrytis vastatrix is an illegitimate superfluous name; Libert published it as a replacement name for Botrytis farinesa. Fr.:Fr. 1029. Therefore the name Bothytis vastatric must be typified by Bothytis farinosa (=Peronospora farinosa), and refers to a different species than Phytophthora Infestans (Ivrt. 7.4).

Distribution: Cosmopolitan.

Substrate: Tubers, leaves, haulms of potato and tomato. Also stems, flowers, fruits, buds on other hosts.

Disease Note: Late blight of potato and tomato. Overwinters in tubers. Several races occur. Also causes leaf blights and, rarely, damping off, flower, and fruit blight in a wide range of hosts.

Host: Principal hosts are Solanaceae including Solanum spp. (potato) and Lycopersicon esculentum (tomato). Also occurs on hosts

in 15 other genera and in ten other families (Erwin & Ribeiro 1996).

Supporting Literature:

Erwin, D.C., and Ribeiro, O.K. 1996. *Phytophthora* Diseases Worldwide. APS Press, St. Paul, Minnesota, 562 pages.

Kroon, L.P.M.M., Bakker, F.I., van den Bosch, B.B.M., Bonants, P.J.M., and Hier, W.B. 2014. Phylogenetic analysis of

Phytophthora species based on millochondrial and nuclear DNS sequences. Fungal Genet. Biol. 41: 766-782.

Puttemans, A. 1936. Reivindicacao visando a denominacae scientifica da doenca da batateira Phytophthora infestans (Mont.)

de Bary. Rodriguesia 2: 341-350

Stamps, D.J. 1983. Phytophthara Infestans. C.M.I. Descript Pathog. Fungi Bact. 838; 1-2 Tucker, C.M. 1991. Tauromy of the gerus Phytophthara de Bary. Univ. Missouri Agric. Esp. Sta. Bull. 153; 1-208

Updated on May 04, 2006

P. infestans is classified in group IV (Stamps et al. 1990). See Tables 4.2 and 4.3 for tabular keys and Appendix 4.9 for a dichotomous key (Ho 1992) in Phytophthora Diseases Worldwide (Erwin and Ribeiro 1996). Morphology is shown in and 4.12G. in Phytophthora Diseases Worldwide (Enwin and Ribeiro 1996). See Fry et al. (1993) and H

*Phytophthora Diseases Worldwide

Link to this site in a senarate window: http://shonanspress.org/42120.html

Sporangia are ovoid, ellipsoid to limoniform, tapering at the base, caduous (pedicel <3 μm), and semipapillate. Average size of sporangia rate under proposo to immorrim, reperting at we case, cast once processing, and sempontage severage see or sporangia ranges from 36 x 22 µm (Tucker 1931), to 29 x 9 µm (Tucker 1931), these dimensions are similar to those of de Bory (1876), Rosenboum (1917), Hockell (1921), K. O. Defor (1928), and Leonan and Green (1929). Sporangophores are compound sympodial (Figure 4.56 in Phytophthora Diesers Worldwide (Erwin and Riberto 1995)) with a small characteristic swelling just below the sporangium (Figures 4.57 x14.46).

Neither hyphal swellings nor chla-chlamydospores with a two layer Lamydospores have been reported, except in a paper from Russia by Patrikeyeva (1979), who noted er wall after incubation for 4 to 9 months on oat-pea agar at 9 to 10°C.

5. Set wyeez

P. Indistant of heterothalic. Anthorodia are amphagnous; ougunia are 31 to 50 pm in diameter (average 38 pm); ouspores formed in place leaves are aplerotic, 24 to 35 pm in diameter (average 30 pm); in artificial culture they measure 24 to 55 pm in diameter. Until the early 1990s, A1 was the only making type found in most of the world, but in central Mexico both A1 and A2 solidates coexisted at the production of the second of the sec a 50:50 ratio (Niederhauser 1991). Inoculation of a leaf with an A2 isolate of P. drechsleri and P. Infectano (A1) induced cosnore. production (Skidmore et al. 1984). Whether or not unique biotypes could arise from unrelated species is unknown. Most like formation of cospores results from stimulation by hormonelike substances emitted by the opposite mating type (see Chapter 3 in Phytophthora Diseases Worldwide (Erwin and Ribeiro 1996).

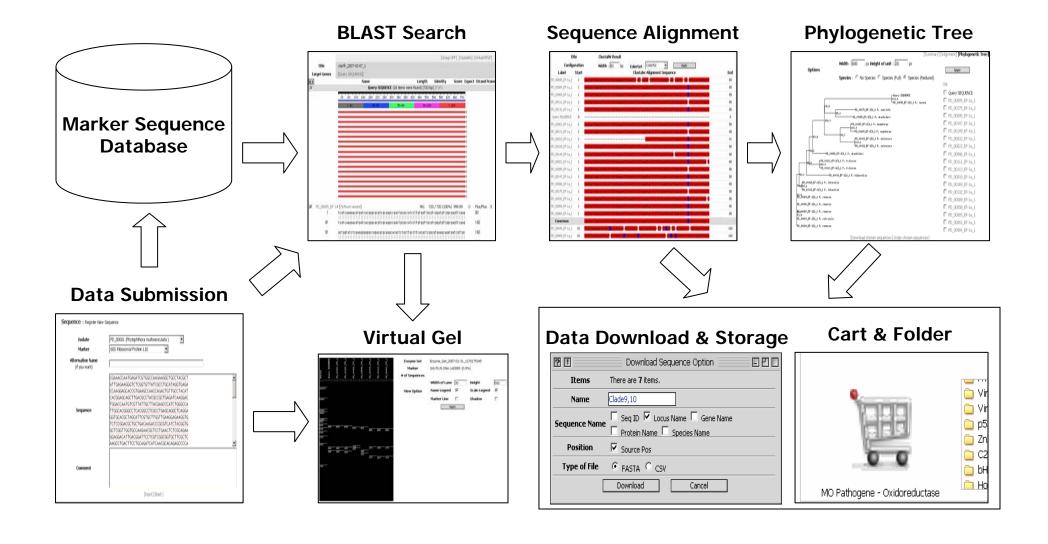
4. Growth Temperatures

The minimum temperature for growth is 4°C, optimum 20°C, and maximum 26°C.

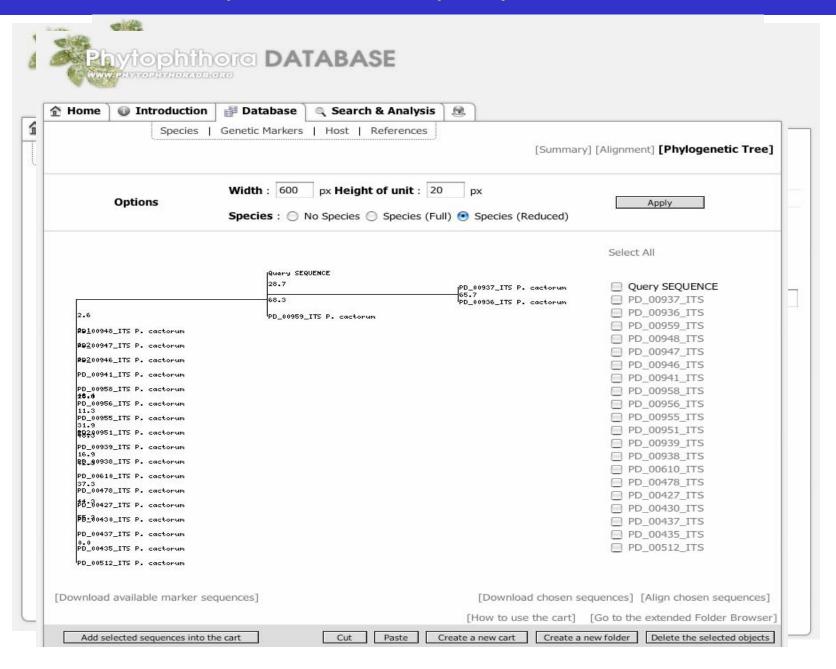
P. infestans affects several hosts in the nightshade family (Solanaceae). It is a severe pathogen of potato (Solanum tuberosum) and tomato (Lycopersicon esculentum) and can lead to 100% crop loss under conditions favorable for disease development and in the



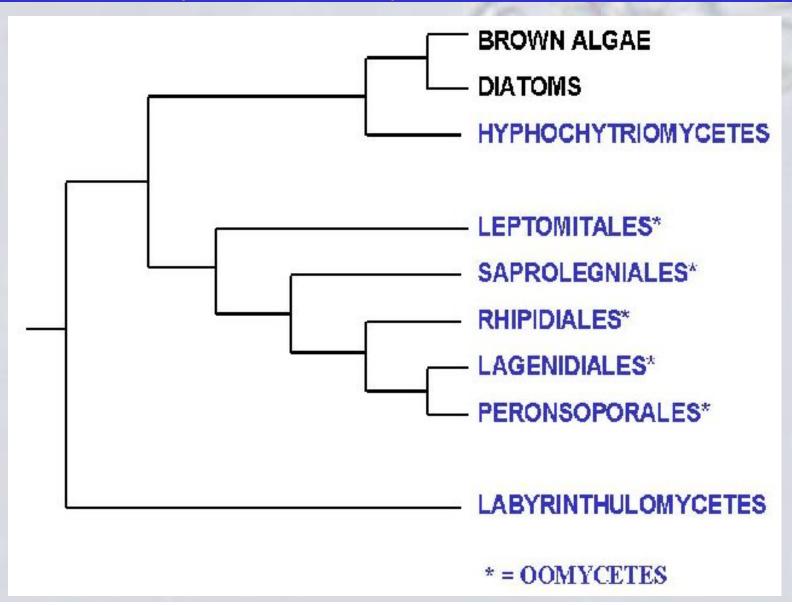
Functionality of the Pytophthora Database

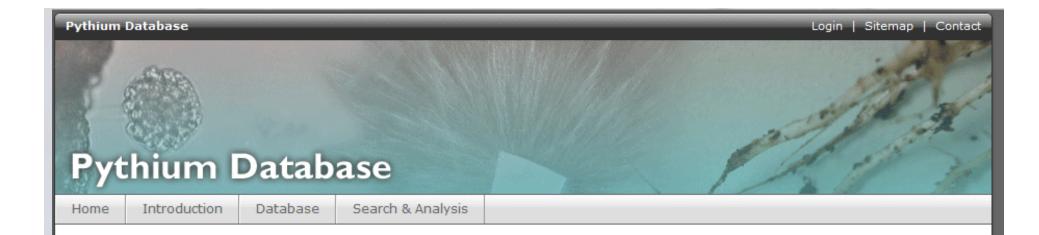


Functionality of the Pytophthora Database



Future Direction Stremenophile/Oomycete Tree of Life?







Pythium Database Login



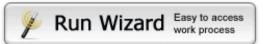
Dear Guest, Welcome to Pythium Database.

- » I want to logout this database.
- » I would like to login the DB as quest



Welcome to Pythium Database

- » About the goals and utility of the database
- » Acknowledgements



- » Data Submission
- » Search the database
- » How to use the database



Pythium Database News



Current statistics of the database

- » 31 Species
- » 364 Isolates
- » 364 Sequences

Link to contributors





Fusarium Database Login

- Dear Guest, Welcome to Fusarium Database.
- » I want to logout this database.
- » I want to modify my login information.
- Welcome to Fusarium Database
- » About the goals and utility of the database
- » Acknowledgements



Fusarium Database News



Current statistics of the database

Link

Link to contributors

- » 15 Species
- » 915 Isolates
- » 2,284 Sequences